

Loading of PureCube NTA or IDA Agarose with nickel

Overview

This protocol describes the loading of PureCube NTA or IDA Agarose with nickel solutions, to obtain Ni-NTA or Ni-IDA Agarose. **Please refer to the appropriate protocol for loading with other transition metals.**

Amounts given in this protocol are for 20 mL NTA or IDA of 50% Agarose suspension, which contains 10 mL agarose. The reaction can be linearly scaled up and down as required.

Please contact us if you have questions or need assistance optimizing a protocol for your application (contact@ www.cube-biotech.com). Additional protocols can also be found at www.cube-biotech/protocols.

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☐ Centrifuge for 50 mL tubes☐ Centrifuge tubes, 50 mL (e.g. Falcon)☐ Vortex mixer☐ End-over-end shaker

Materials

PureCube NTA Agarose (10 mL, Cube
Biotech #31703) or PureCube IDA Agarose
(10 mL, Cube Biotech #30703)
Nickel II sulfate
Sodium chloride
Sodium acetate trihydrate
Tris base
Acetic acid
Ethanol
Hydrochloric acid

Loading_Ni-NTA or Ni-IDA_Ag_1605.1

Solutions and buffers

Sodium acetate buffer, pH 6.0, 100 mL

Component	Final concentration	Molecular weight (g/mol)	Stock concentration	Amount needed for buffer
Sodium acetate trihydrate	50 mM	136.08	n.a.	680 mg

Instructions: Dissolve sodium acetate in 80 mL water, adjust the pH to 6.0 with acetic acid. Add water to a total volume of 100 mL.

Nickel sulfate solution, 50 mL

Component	Final concentration			Amount needed for buffer
Nickel II sulfate hexahydrate	2.5% (w/v)			1.25 g
Instructions: Dissolve in 50 ml water.				

Wash Buffer, 100 mL

Component	Final concentration	Molecular weight (g/mol)	Stock concentration	Amount needed for buffer
Sodium chloride	150 mM	58.44	n.a.	877 mg
Acetic acid	200 mM	60.05 Density 1.05 g/mL		1.14 mL

Instructions: Dissolve sodium chloride in 80 mL water, then add acetic acid. Add water to a total volume of 100 mL.

Tris buffer, pH 7.5, 200 mL

Component	Final concentration	Molecular weight (g/mol)	Stock concentration	Amount needed for buffer
Tris base	20 mM	121.14		484 mg

Instructions: Dissolve Tris base in 160 mL water, adjust the pH to 7.5 with hydrochloric acid. Add water to a total volume of 200 mL.

Agarose Storage Buffer, pH 6.5, 50 mL

Component	Final concentration	Molecular weight (g/mol)	Stock concentration	Amount needed for buffer
Sodium acetate trihydrate	20 mM	136.08	n.a.	135 mg
Ethanol	20 % (v/v)		100 % (v/v)	10.2 mL

Instructions: Dissolve sodium acetate in 30 mL water, adjust the pH to 6.5 with acetic acid. Add 9.6 mL water and 10.2 mL ethanol to yield a total volume of 50 mL.

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Procedure

- Transfer 20 mL PureCube NTA or IDA Agarose suspension into a 50 mL centrifuge tube.
- 2. Spin the tube for 5 min at 500 x g to pellet the agarose. Remove the supernatant. Resuspend with 20 mL double distilled water.
- 3. Wash two more times with 20 mL water.
- 4. Wash 3x with 20 mL 50 mM sodium acetate, pH 6.0.
- 5. Wash 1x with 20 mL double distilled water.
- 6. Add 20 ml 2.5% nickel sulfate solution and incubate for 2 h.
- 7. Wash 4x with 20 mL double distilled water.
- 8. Add 20 mL Wash Buffer and incubate for 10 min.
- 9. Wash 1x with 20 mL double distilled water.
- 10. Wash 6x with 20 mL 20 mM Tris-HCl, pH 7.5.
- 11. Wash 1x with 20 mL double distilled water.
- 12. Resuspend the Ni-NTA or Ni-IDA Agarose in 20 mL Agarose Storage buffer, yielding a 50% suspension. Store at 4°C.

Tip: The loading reaction can be scaled up and down linearly, by increasing or decreasing the amounts of buffers and solutions described in this protocol.

